

1 Organic Adsorption onto Iron Hydroxide and Sulfide Minerals: 2 Implications for Ceres Sample Return Analysis

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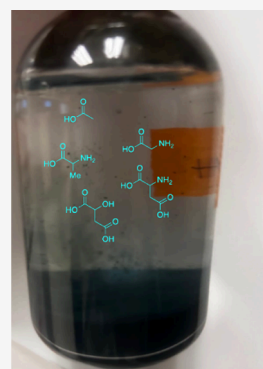
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6 **ABSTRACT:** Characterizing the past and present habitability potential of Ceres is an important goal of
7 a future NASA mission to the dwarf planet. Signs of organic molecules (potentially amino acids,
8 carboxylic acids, etc.) and their potential interactions with surrounding minerals may be present on the
9 surface of Ceres. In addition, iron minerals are postulated to be present based on Dawn's gamma ray and
10 neutron detector measurements, and iron minerals are known to be able to adsorb organic molecules.
11 To understand if these iron minerals could adsorb prebiotically relevant organics on the surface of Ceres,
12 we conducted an experimental study to test the adsorption of a set of simple organics (alanine, glycine,
13 acetate, malic acid, and aspartic acid) on two classes of iron minerals found in carbonaceous chondrites
14 (Fe-sulfides and Fe-hydroxides, synthesized via coprecipitation). Our experiments revealed little to no
15 adsorption of amino acids onto each of the Fe-minerals. Our results also revealed that the redox state of
16 the Fe-hydroxide minerals remained stable throughout the experiment; however, Fe-sulfide precipitates
17 exhibited reduction of Fe(III) to Fe(II). Additionally, there was negligible adsorption of glycine onto
18 mixed-valent iron mineral systems in a synthetic brine solution. This work shows that organic acids
19 remain stable in the presence of Fe-bearing minerals and are mobile when dried and rehydrated by
20 brines, meaning that these organics may be stable on Ceres' surface but could be further mobilized during cryovolcanic events, the
21 products of which may be used for further reactivity toward other complex molecules with astrobiological relevance.

22 **KEYWORDS:** *Ceres, organic adsorption, iron sulfides, iron (Hydr)oxides, amino acids, habitability*



23 ■ INTRODUCTION

24 Ceres, a dwarf planet located within the asteroid belt, has
25 become a prime target of astrobiological significance, especially
26 because its abundant water-ice content, geochemically rich
27 past, and presence of organic compounds suggest that it could
28 have been habitable in the past and possibly at present.^{1–5} The
29 2023–2032 Planetary Science and Astrobiology decadal survey
30 has included a sample return mission to Ceres as a part of the
31 New Frontiers mission portfolio.⁶ That mission would address
32 (1) Ceres' current habitability potential and its contribution to
33 studying the habitability of icy worlds; (2) identify Ceres'
34 origins; and (3) quantify the evolution of organic compounds
35 in a long-lived brine reservoir, with implications for under-
36 standing other inner solar system planetary bodies.^{4–8}
37 Observations by Dawn's visible and infrared mapping
38 spectrometers (VIR) reveal the presence of Mg/Al phyllosil-
39 licates, ammoniated phyllosilicates, carbonates, and Fe-bearing
40 minerals (magnetite and Fe-sulfides) believed to be from
41 aqueous alteration.^{5,7,9,10} Additionally, modeling efforts
42 revealed that the darkening agents found on Ceres' surface
43 are likely to be magnetite and/or Fe sulfide minerals with
44 amorphous organics, which are consistent with the data
45 obtained from the Dawn gamma ray and neutron detector
46 (GRaND).^{11,12} Dawn also revealed the presence of aliphatic

organics at Ernutet crater (based on the 3.4 μm band detected
47 by VIR) similar to those found on carbonaceous chondrite
48 meteorites and likely endogenously sourced.^{1,13,14} As one of
49 the largest icy planetesimals, Ceres' evolution as a result of
50 water-rock alteration may help us to understand prebiotic
51 chemistry in bodies that contributed water and organics to the
52 inner solar system.⁵

53 Iron minerals are highly reactive and can be easily oxidized
54 or reduced by their surrounding environmental conditions as
55 well as by organic reagents.^{15,16} Iron minerals have a range of
56 oxidation states, each state with unique structural and reactive
57 properties that can influence the fate of organic molecules and
58 can shield organics from photochemical and biological
59 oxidation/degradation.^{17–19} Ceres' inferred alkaline condi-
60 tions²⁰ could enable the precipitation of reactive Fe-hydroxides
61 and Fe-sulfides that could also react with organic molecules. 62

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63 Mineral surfaces can serve as a binding site for organic or
64 inorganic compounds to adhere to the mineral surface via
65 different adsorption mechanisms (e.g., ligand exchange, H-
66 bonding, Lewis acid–base interactions, etc.).^{16,21,22}

67 To understand the complex chemistry that may have
68 occurred on the surface of Ceres to contextualize its potential
69 habitability, it is important to characterize the interactions
70 between precipitated Fe minerals and a potential organic
71 material. The characterization of the organic material on Ceres
72 is currently limited to information from the structure of the 3.4
73 μm VIR band that indicates the presence of methyl (CH_3) and
74 methylene (CH_2) functional groups.^{23,24} The abiotic synthesis
75 of organic molecules (such as amino acids and α -hydroxy
76 acids) on planetary bodies is of particular interest given that
77 they are critical to life as we know it, for example, amino acids
78 are necessary for protein generation,²⁵ while α -hydroxy acids
79 are key intermediates in the tricarboxylic acid cycle which is
80 found in many organisms.^{26,27} Like carbonaceous chondrites
81 (CC chondrites), Ceres could have locales for abiotic amino
82 and α -hydroxy acid synthesis via mechanisms such as Strecker
83 synthesis, reductive amination, and aldehyde plus ammonia
84 reactions.^{28–30} Given that Ceres has reactive minerals as well
85 as organic molecules to drive organic acid synthesis, we
86 investigated if organic acid adsorption occurs onto minerals
87 found on Ceres once they have been synthesized.

88 Here, we tested several Fe-hydroxide and Fe-sulfide mineral
89 types, which are known to transform into magnetite (and other
90 iron oxides), making them relevant to the GRaND measure-
91 ments.^{31,32} We investigated the effect of the redox state of the
92 iron mineral on the adsorption of five organic molecules
93 (alanine, glycine, acetate, malic acid, and aspartic acid) of
94 prebiotic relevance and have been shown to be synthesized
95 abiotically.^{28–30} In fact, reactions with iron minerals have been
96 demonstrated to produce abiotic amino acids, α -hydroxy acids,
97 and others via iron minerals, including Fe-hydroxides and Fe-
98 sulfides in our group and by other investigators.^{29,33–35} We
99 also investigated whether these organics are mobile if the
100 mineral/organic mixture is desiccated and rehydrated with
101 brine, such as what might occur in successive brine flows in
102 cryovolcanic sites on Ceres.

103 ■ MATERIALS AND METHODS

104 All experiments were performed at room temperature in a
105 nitrogen-filled glovebox. No adjustment was made to the
106 temperature, as we were focused on exploring the mineral–
107 organic interaction. Future work could explore varying the
108 temperature. All solutions were prepared with deionized Milli-
109 Q water (18.2 $\text{M}\Omega\cdot\text{cm}$) that had been sparged with N_2 (30
110 min per 100 mL H_2O). In order to explore these reactions, the
111 iron minerals were synthesized within the lab based on
112 previously published literature.^{17,33} Iron solutions for mineral
113 precipitation were prepared using $\text{FeCl}_2\cdot 4\text{H}_2\text{O}$ and/or
114 $\text{FeCl}_3\cdot 7\text{H}_2\text{O}$ (Fisher Scientific), sulfide solutions were
115 prepared using $\text{Na}_2\text{S}\cdot 9\text{H}_2\text{O}$ (Sigma-Aldrich), and organic
116 acid solutions were prepared using the following: L-alanine
117 (99% pure, Thermo Scientific), glycine (98% pure, Acros
118 Organics), sodium acetate (99% pure, Fisher Chemical), DL-
119 malic acid (99–100.5% pure, VWR), and aspartic acid (99%
120 pure, Sigma). The organics were used without any additional
121 purification. Titrating solutions were prepared by using NaOH
122 (Avantor) and HCl (Sigma-Aldrich).

123 **Organic Acid Adsorption Experiments in Aqueous**
124 **Solution.** In our experiments, we refer to adsorption as the

surface interaction between our Fe minerals and organic acids, 125
primarily because our minerals can undergo Lewis acid–base 126
interactions rather than absorption, given the lack of a layered 127
structure (e.g., clay minerals) in our minerals. The sorption of 128
organics (alanine, glycine, acetate, malic acid, aspartic acid) 129
onto lab-synthesized Fe-hydroxide and Fe-sulfide minerals was 130
tested at pH 10 (Table SI-1). The organic acid adsorption 131
experiments were performed at 25 °C and at standard 132
atmospheric pressure (1 atm) to understand adsorption 133
under stable conditions and minimize the introduction of 134
additional variables. This method allowed us to isolate the 135
effects of adsorption on a stable temperature and also assess 136
the influence of temperature on our Ceres brine freeze-dried 137
experiments (described below). A 140 mL stock solution of 138
Fe-hydroxide was prepared by precipitating 131.25 mL of 139
dissolved $\text{FeCl}_2\cdot 4\text{H}_2\text{O}$ and/or $\text{FeCl}_3\cdot 7\text{H}_2\text{O}$ with 8.75 mL of 140
1 M NaOH. A 160 mL stock solution of FeS was prepared by 141
combining 80 mL of dissolved $\text{FeCl}_2\cdot 4\text{H}_2\text{O}$ and/or 142
 $\text{FeCl}_3\cdot 7\text{H}_2\text{O}$ with 80 mL of $\text{Na}_2\text{S}\cdot 9\text{H}_2\text{O}$. Twenty-five mM 143
organic stock solutions were prepared and titrated to pH 10 144
with NaOH or Na_2S . For each experiment, 20 mL of 145
precipitated mineral slurry and 5 mL of organic stock solution 146
were combined to make a solution with a final Fe 147
concentration of 50 mM, NaOH or $\text{Na}_2\text{S}\cdot 9\text{H}_2\text{O}$, and 5 mM 148
organic stock solution while being continuously stirred. For 149
experiments using Fe-hydroxides, pH values of the organic and 150
the Fe mixtures were below 10; therefore, additional NaOH 151
was added to reach a pH value of 10. High pH was needed to 152
create the mineral; however, this is relevant to possible water- 153
rock interactions on Ceres. Processes such as serpentinization 154
could be a source of magnetite, raise the pH of surrounding 155
fluids (pH > 9), and create reducing conditions.^{28,29} The pH 156
prior to adjustment for experiments with 100% Fe^{2+} was ~6– 157
7, and experiments with 50% = $\text{Fe}^{2+}:\text{Fe}^{3+}$ and 100% Fe^{3+} were 158
~1.50 – 2.0. This solution was then titrated to pH 10 using aq. 159
NaOH and aq. HCl. The effect of the mineral's redox state on 160
organic sorption was tested by varying the $\text{Fe}^{2+}:\text{Fe}^{3+}$ ratio. 161
Three iron ratios ($\text{Fe}^{2+} = 100\%$, $\text{Fe}^{2+}:\text{Fe}^{3+} = 50\%$, and $\text{Fe}^{3+} = 162$
100%) were tested for each type of mineral. Each experiment 163
was placed in a stir plate and was constantly stirred for 2 weeks 164
at 100 rpm. Each experiment was performed four times. A full 165
list of experiments (Table SI-1) and images of the experimental 166
serum vials (Figures SI-1 and SI-2) can be found in the 167
Supporting Information. Six 1 mL aliquots were taken from 168
each mineral/organic solution at $t = 0$ (after titrating to pH 169
10), 1 week, and 2 weeks. The pH of the experiments was 170
measured using a Thermo Scientific Orion Star A211 171
Benchtop pH meter at each sampling time point (Figure SI- 172
23). The liquid phase was analyzed with ^1H NMR to 173
determine organic concentration, and the solid material was 174
analyzed with iron colorimetry to determine Fe redox state. At 175
the end of each experiment, the remaining solution and 176
minerals were removed from the stir plate and left to settle 177
inside the glovebox to prevent atmospheric oxidation of our Fe 178
minerals. The supernatant was then pipetted out, and the 179
bottle was covered with parafilm and transferred to an SP 180
Scientific Virtis AdVantage benchtop freeze-dryer to freeze-dry 181
over 3 days. The benchtop freeze-dryer was not equipped with 182
an inert gas supply. All experiments were removed from the 183
glovebox and were exposed to atmospheric oxidation during 184
the freeze-dry processes, which may have altered the redox 185
state of the minerals at the end of the experiment. Once dry, 186

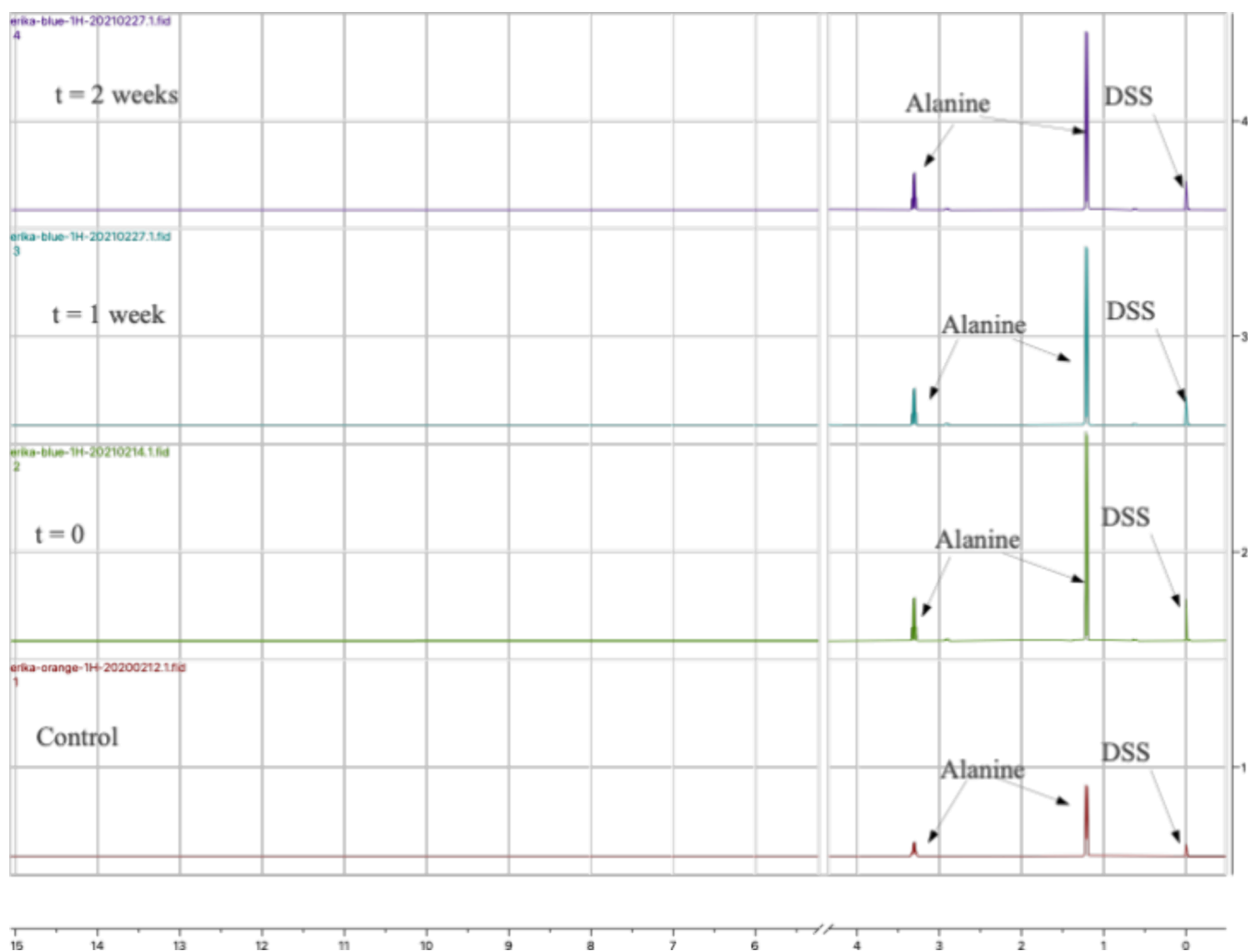


Figure 1. NMR spectra of alanine peaks from experiment containing Fe(II)-hydroxide at pH 10. The water peak is omitted for the sake of clarity.

187 the samples were transferred to microcentrifuge tubes and
188 stored in the glovebox until ready for further analysis.

189 **Ceres Brine Rehydration Experiments.** To estimate the
190 composition of a Ceres brine (representing eruption from near
191 surface brine reservoirs onto the surface), we modeled in
192 Geochemist's Workbench a brine composition for a pH of 6.5
193 and water/rock ratio of 10. The GWB output (Table SI-9)
194 derived the molal concentration of major aqueous species in a
195 brine; we chose to include in our estimates those species that
196 were present at concentrations above 0.001 molal (these
197 included HCO_3^- , NH_4^+ , Na^+ , Cl^- , Ni_2^+ , $\text{NaHCO}_3(\text{aq})$,
198 MgHCO_3^+ , Mg_2^+ , and K^+). We derived a recipe for a
199 laboratory Ceres brine solution (Table SI-9) including salts
200 of these species to approximate the concentrations from Table
201 SI-8. The Ceres brine analogue solution was composed of 7.86
202 $\times 10^{-2}$ M $(\text{NH}_4)_2\text{CO}_3$ (Sigma-Aldrich), 1.66×10^{-2} M
203 NaHCO_3 (Fisher Scientific), 1.91×10^{-2} M NH_4Cl (Fisher
204 Scientific), 6.80×10^{-3} M $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$ (Fisher Scientific),
205 1.71×10^{-2} M NiCl_2 (Sigma-Aldrich), 6.88×10^{-3} M MgCO_3
206 (Sigma-Aldrich), and 2.21×10^{-3} M KCl (Mallinckrodt);
207 titrated to pH 6.5. Stock solutions of Fe-hydroxide and Fe-
208 sulfide minerals for the Ceres brine experiments were prepared
209 in the same manner as that stated in the aqueous organic acid
210 adsorption experiments. For these brine experiments, however,

only one redox state (50% = $\text{Fe}^{2+}:\text{Fe}^{3+}$) and only one organic
molecule (glycine) were tested.

Control experiments of Fe-hydroxide (50% = $\text{Fe}^{2+}:\text{Fe}^{3+}$ and
NaOH) and Fe-sulfide (50% = $\text{Fe}^{2+}:\text{Fe}^{3+}$ and Na_2S) were
performed by precipitating 50 mL of each mineral without the
addition of any organics and freeze-dried without the removal
of any supernatant. A thermocouple was inserted into each
bottle to monitor the conditions of the mineral throughout the
freezing/drying process. A Kimwipe was used to cover the
thermocouple and avoid cross-contamination between experi-
ments. A freeze-dryer recipe was developed to ensure that the
samples would dry in a period of 3 days. A detailed description
of the freeze-drying process is provided in the SI, involving
only one freezing cycle followed by drying. Once dry, the
minerals were removed and brought into the glovebox, where
they were rehydrated with 50 mL of the Ceres brine solution.
Mineral/brine solutions were placed on a stir plate at 100 rpm
for 2 weeks, with sampling in between. Six 1 mL aliquots were
taken prior to freeze-drying, upon addition of the Ceres brine
solution ($t = 0$), and at 1 and 2 weeks of stirring post Ceres
brine solution addition. These samples were frozen to preserve
them for later analysis with nuclear magnetic resonance
(NMR) and iron colorimetry. These experiments were then
repeated but with the addition of glycine, which was weighed

235 out and added directly to the mineral before freeze-drying. All
236 experiments were repeated three times.

237 **Analytical Techniques.** ¹H Nuclear Magnetic Resonance
238 (NMR) Spectroscopy. Prep for NMR analysis involved
239 centrifuging three of the samples, transferring 0.75 mL of
240 supernatant to a new microcentrifuge tube with 0.25 mL of 1
241 M NaOH to precipitate out any soluble Fe, and centrifuging
242 again. The supernatant was then used to prepare NMR
243 samples by taking 0.54 mL from the microcentrifuge tube and
244 adding 60 μL of DSS in D₂O (sodium trimethylsilylpropane-
245 sulfonate/deuterium oxide) as standard to an NMR tube. ¹H
246 NMR spectra were acquired on a 400 MHz Bruker
247 spectrometer equipped with an auto sampler. The spectra
248 were processed with the MestReNova NMR analysis software.
249 The concentrations (mM) of each organic in the mineral
250 supernatant were also calculated by using the NMR methyl
251 peak areas in comparison to the DSS peak (which was
252 integrated to 1). The NMR spectra can be found in the SI for
253 each experiment. The average percent change was calculated
254 by using the following equation, percent increase or decrease
255 (%) = (t₀ - t_x)/t₀ where t₀ is the concentration of organic
256 detected in the supernatant at t = 0 and t_x is the concentration
257 detected in the supernatant of samples taken at either t = 1
258 week or t = 2 weeks. Sample t = 0 is the time point at which
259 the organic acids were added to the experiment and processed
260 for NMR (within 10 min).

261 **Fe²⁺ Colorimetry.** To track the oxidation state of iron, 1 mL
262 of 10 M HCl was added to three liquid–solid samples that
263 were then heated at 100 °C for a few minutes to dissolve the
264 mineral precipitate. Once fully dissolved, the samples were
265 diluted and run in a Thermo Fisher GENESYS 30 Visible
266 spectrophotometer set to a wavelength of 510 nm using a
267 slightly modified version of a Fe²⁺ colorimetry technique.³⁶
268 The % of Fe²⁺ in the dissolved samples was calculated, and the
269 averages from all repeats are plotted. The ratio of Fe²⁺/Fe(T)
270 was used to report the percent of Fe(II) in our experiments,
271 where FeT (Fe Total) is equal to the sum of the Fe²⁺ and Fe³⁺.

272 **Visible Near-Infrared (VNIR) Reflectance Spectroscopy.**
273 The spectral reflectance of lyophilized mineral samples at 2
274 weeks was obtained by an analytical spectral device (ASD)
275 FieldSpec3 spectroradiometer with a wavelength range of 0.3–
276 2.5 μm and a resolution of 3 (350–1000 nm) or 10 nm
277 (1000–2500 nm). Sample spectra were collected against a
278 Spectralon white reference (Figure 1). The dried mineral
279 samples were placed on either white weighing paper (VWR) or
280 black Cinefoil and illuminated by a fiber optic cable with a
281 while light source (quartz tungsten halogen) on a hand-held
282 contact probe. The powdered samples were pressed against the
283 contact probe, and the spectral reflectance was collected on
284 RS.³ In between samples, the probe was wiped with a Kimwipe.
285 The samples were collected and saved post VNIR analysis.

286 ■ RESULTS AND DISCUSSION

287 **Organic Acid Adsorption onto Fe (Oxy)hydroxides.** In
288 Fe-hydroxide experiments, we observed similar trends of the
289 organic acids' (Table 1) adsorption onto each of the Fe-
290 hydroxide mineral conditions. In experiments containing 100%
291 Fe(II)-hydroxide minerals, adsorption of all organic com-
292 pounds tested increased to approximately <16% at t = 2 weeks.
293 The two most adsorbed organic acids in the Fe(II)-hydroxide
294 experiments were malic acid and aspartic acid, with each
295 adsorbing 14.25% and 15.85%, respectively (Figure 2A). In
296 experiments with 50% = Fe(II)/Fe(III)-hydroxide the percent

Table 1. Functional Groups and Properties of Organics Considered in This Work

organic	functional Group	pK _a	reference
alanine	carboxyl (COOH), amine (NH ₂)	pK ₁ = 2.3, pK ₂ = 9.8	37
glycine	carboxyl (COOH), amine (NH ₂), H atom	pK ₁ = 2.3, pK ₂ = 9.6	38
acetate	carboxyl (COOH), ester	pK _a = 4.76	39
malic acid	dicarboxyl (COOH)	pK ₁ = 3.4, pK ₂ = 5.2	40
aspartic acid	carboxyl (COOH), amine (NH ₂)	pK _{a1} = 2.0, pK _{a2} = 3.9, pK _{a3} = 9.9	41

297 adsorbed remained relatively constant near ~0% adsorbed at 1
298 and 2 weeks (Figure 2B). In experiments with 100% Fe(III)-
299 hydroxide, all of the organic acids desorbed at 1 week and were
300 later re-adsorbed except alanine and glycine (Figure 2C).

301 The pH decreased for experiments beginning with 100%
302 Fe(II) and with 50% = Fe(II)/Fe(III), while the pH increased
303 for experiments beginning with 100% Fe(III) (Figure SI-23).
304 The decrease in pH followed the same trend for each mineral
305 type; for example, in experiments with 100% Fe(II)-hydroxide
306 precipitates, there was a decrease at t = 1 week and a slight
307 increase at t = 2 weeks. The pH of the experiment between
308 Fe(III)-hydroxides and the presence of alanine and glycine in
309 solution remained stable at a pH level of 9.5, whereas all other
310 organic molecules experienced lower pH's throughout the
311 course of the experiment. The 50% = Fe(II)/Fe(III)-hydroxide
312 experiments experienced a similar trend, where the experi-
313 ments decreased from a pH of 10 to below 9.0 (Figure SI-23).
314 Lastly, the pH of the experiments with 100% Fe(III)-hydroxide
315 increased, with some increasing to approximately ~10.3 over
316 the 2 week time. The colorimetry results revealed that the
317 redox of Fe hydroxide minerals remained stable throughout the
318 experiments. The experiments beginning with 100% Fe(II)-
319 hydroxide minerals remained reduced with the Fe²⁺/FeT
320 nearly at or near 100% (Figure 3A). In experiments beginning
321 with 50% = Fe(II)/Fe(III)-hydroxides, the minerals remained
322 near their original redox state at 50% Fe²⁺ and 50% Fe³⁺
323 (Figure 3B). Lastly, in the experiments beginning with 100%
324 Fe(III)-hydroxides, the minerals remained oxidized with Fe²⁺/
325 FeT being less than 5% (Figure 3C).

326 **Organic Acid Adsorption onto Fe Sulfide Minerals.** In
327 Fe-sulfide mineral experiments, we did not observe substantial
328 organic acid adsorption. In 100% Fe(II)-sulfide experiments, a
329 trend was observed with alanine, malic acid, and aspartic acid,
330 where organic acid concentrations remained the same for t = 0
331 and t = 1 week and were later desorbed around 2 weeks
332 (Figure 2D). For experiments with 100% Fe(II)-sulfide
333 minerals, glycine showed the highest adsorption (~19.15%)
334 at 1 week, followed by desorption, for a total of 2.22%
335 adsorbed at 2 weeks. All organic acids reacting with 100%
336 Fe(II)-sulfides, except for glycine, showed less than 5%
337 adsorption for 1 week and 2 weeks. In Fe-sulfide mineral
338 experiments starting with 50% = Fe(II)/Fe(III), malic acid was
339 the highest adsorbed organic compound at 1 week (~8.16%)
340 and adsorption increased to 10.61% at 2 weeks. In the 50% =
341 Fe(II)/Fe(III)-sulfide experiments, less than 5.00% of alanine,
342 acetate, and aspartic acid were adsorbed in 1 week, followed by
343 adsorption increasing at 2 weeks for a total of 6.10, 15.00, and
344 12.34% being adsorbed, respectively. Lastly, the experiment
345 with 50% = Fe(II)/Fe(III) and glycine showed no adsorption
346 until 2 weeks, where 9.32% of glycine was adsorbed (Figure
347 2E). In experiments with 100% Fe(III)-hydroxides reacting

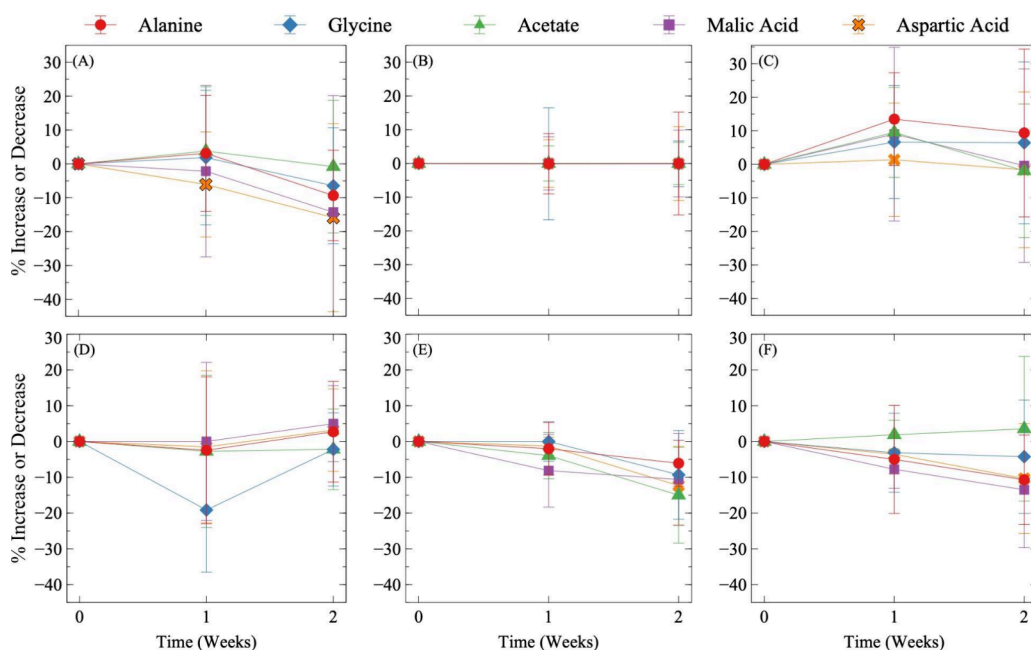


Figure 2. Average percent change of organic compounds in solution at $t = 1$ week and 2 weeks compared to $t = 0$ for both Fe (oxy)hydroxide (A–C) and Fe sulfide (D–F) minerals. Percent change greater than 0 (positive values) represents organics released from the mineral (a higher concentration of organic was detected in the supernatant). Percent less than 0 (negative values) represents the percentage of organics bound to the mineral increase at $t = 1$ week and $t = 2$ weeks compared to $t = 0$. (A) Experiments with 100% Fe^{2+} and NaOH, (B) experiments with 50% $\text{Fe}^{2+}:\text{Fe}^{3+}$ and NaOH, (C) experiments with 100% Fe^{3+} and NaOH, (D) experiments with 100% Fe^{2+} and Na_2S , experiments with 50% $\text{Fe}^{2+}:\text{Fe}^{3+}$ and Na_2S , and (F) experiments with 100% Fe^{3+} and Na_2S . The error bar represents the standard deviation from the average of experiments repeated four times.

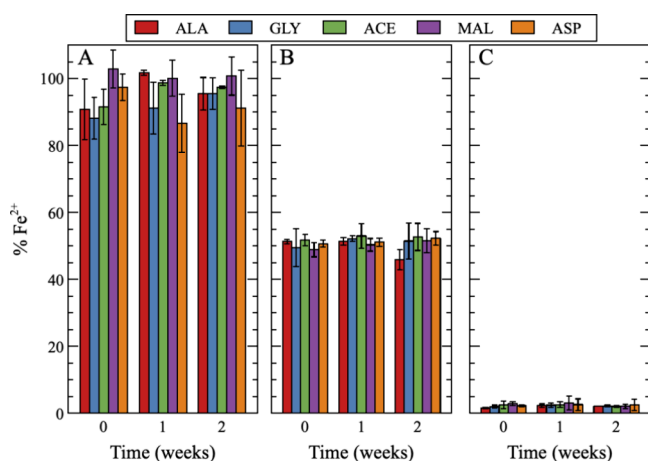


Figure 3. Fe oxidation state of Fe-hydroxide minerals reacted with the organic acids. (A) Experiments with Fe(II)-hydroxide and organic acids, (B) experiments with 50% = Fe(II)/Fe(III)-hydroxides reacting with organic acids, and (C) experiments 100% Fe(III)-hydroxide reacting with organic acids. The error bar is the standard deviation of the averages from all repeats.

experiment (Figure 4A). In experiments with 50% = Fe(II)/Fe(III)-sulfide minerals, the partial Fe(III) in our minerals was 358 f4 359

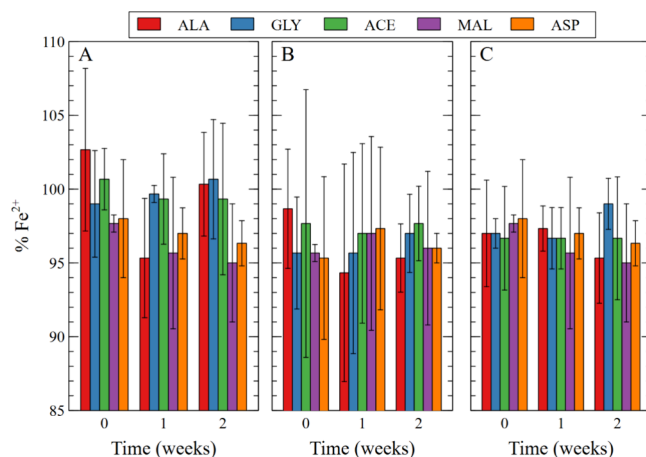


Figure 4. Fe oxidation state of Fe-sulfide minerals reacted with the organic acids. (A) Experiments with Fe(II)-sulfide and organic acids, (B) experiments with 50% = Fe(II)/Fe(III)-sulfide reacting with organic acids, and (C) experiments with 100% Fe(III)-sulfide reacting with organic acids. The error bar is the standard deviation of the averages from all repeats.

348 with the organic molecules, minor adsorption was observed
349 with alanine (5.01%), glycine (3.15%), malic acid (7.77%), and
350 aspartic acid (3.59%) at 1 week and increased to 10.70, 4.26,
351 13.49, and 10.36% at 2 weeks, respectively (Figure 2E).

352 The Fe in the Fe-sulfide minerals was redox sensitive during
353 the precipitation of our minerals. The precipitate formed for all
354 starting Fe solutions was a black material despite the original
355 oxidation state of the salt (i.e., FeCl_2 and FeCl_3). The
356 colorimetry results of our Fe(II)-sulfide minerals show that the
357 Fe remained as Fe(II) throughout the duration of the

reduced to Fe(II) during the initial sampling at $t = 0$ and 360
remained reduced ($>95.00\%$ $\text{Fe}^{2+}/\text{FeT}$) for the 2 weeks 361
(Figure 4B). The Fe(III)-hydroxides were also reduced during 362
the start of the experiments ($t = 0$) and remained reduced 363
($>95.00\%$ $\text{Fe}^{2+}/\text{FeT}$) throughout the 2 weeks of the experi- 364
ment (Figure 4C). 365

366 **Ceres Brine Solution Experiments.** Adsorption of
367 glycine was tested onto 50% = Fe(II)/Fe(III)-hydroxide and
368 50% = Fe(II)/Fe(III)-sulfide minerals. ^1H NMR was
369 performed on the supernatant samples in solutions prior to
370 freeze-drying, after freeze-drying, and rehydrating with the
371 Ceres brine solution ($t = 0$), after constantly stirring that
372 solution for 1 and 2 weeks. The spectra of the controls showed
373 only the DSS/D₂O standard peaks and the glycine peak after it
374 was added. For the Fe-50% = Fe(II)/Fe(III)-hydroxide and
375 50% = Fe(II)/Fe(III)-sulfide experiments without organic
376 acids, there were no additional peaks detected. In the 50% =
377 Fe(II)/Fe(III)-hydroxide, there was $\sim 6.00\%$ adsorbed glycine
378 and as later released at 2 weeks (Figure 5). In contrast, the

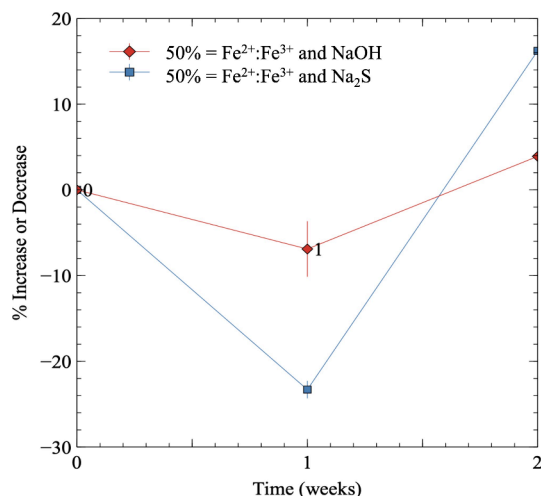


Figure 5. Average percent increase/decrease of glycine in 50% = Fe(II)/Fe(III)-hydroxide and 50% = Fe(II)/Fe(III)-sulfide minerals in a Ceres brine solution. Positive values represent organic molecules released from the mineral (a higher concentration of organic molecules detected in the supernatant). Negative values represent the percentage of organic molecules bound to the mineral increase at $t = 1$ week and $t = 2$ weeks compared to $t = 0$. The error bars are the standard deviation of the three experimental repeats.

379 50% = Fe(II)/Fe(III)-sulfide experiment showed that $\sim 20\%$ of
380 glycine was adsorbed to the mineral at 1 week and was later
381 released at 2 weeks (Figure 5). Some differences between this
382 work and the previous work could be attributed to mineral
383 alterations (e.g., oxidation) in freeze-drying and other
384 processing.

385 **Reflectance spectroscopy.** A range of overlapping
386 electronic absorptions in the $0.35\text{--}2.5\ \mu\text{m}$ wavelength range
387 can be caused by the presence of ferrous and/or ferric Fe via
388 intervalence charge transfer, oxygen–metal charge transfer, and
389 crystal field splitting.^{42–45} We observed two broad classes of
390 mineral spectra. Our 100% Fe(II)-hydroxide minerals may be a
391 chloride green rust, and it is important to note that the
392 resulting oxidation was a result of atmospheric oxidation when
393 the samples were removed from the glovebox for analysis. We
394 have compared the reflectance spectra obtained from our
395 experiments to the reflectance spectra of minerals in the
396 RELAB VNIR library database.⁴⁶

397 First, in Fe-hydroxide experiments with 100% Fe(II) (Figure
398 6A) and 100% Fe(III) (Figure 6C), there are strong electronic
399 absorptions from 0.4 to 0.55 and 0.9 μm , consistent with
400 abundant Fe oxides, possibly hematite (Figure 7). In the 100%
401 Fe(II)-hydroxide experiments (Figure 6A), some samples

(acetate and to a lesser extent alanine, glycine, and malic
402 acid) exhibited hydration features at $1.9\ \mu\text{m}$ associated with
403 either structural or adsorbed water, suggesting the presence of
404 another phase either with water in its mineral structure or a
405 phase which is hygroscopic and adsorbs water to its surface
406 easily. The hydration features are even stronger in the 100%
407 Fe(III)-hydroxide experiments (Figure 6C), where all samples
408 have 1.4 and $1.9\ \mu\text{m}$ absorptions, indicating hydration of the
409 sample. A small shoulder near $2.3\ \mu\text{m}$ in the 100% Fe(III)-
410 hydroxide experiments (Figure 6C) is a metal–OH vibration.
411 The asymmetrical shape of the feature is more likely an Fe-
412 oxide phase such as goethite, akageneite, butlerite, or
413 ferrihydrite, which can also show small, asymmetrical
414 absorptions in this area.^{47,48} We note that the control in the
415 100% Fe(II)-hydroxide (Figure 6A) experiment is more
416 consistent with magnetite, which may have undergone
417 atmospheric oxidation while we transferred the samples to
418 the freeze-dryer (see below).
419

Second, in Fe-hydroxide experiments with 50% = Fe²⁺:Fe³⁺
420 (Figure 6B) and all Fe-sulfide experiments (Figure 6D–F), we
421 see spectra broadly consistent with magnetite. Magnetite is a
422 mixed valence iron oxide whose structure contains abundant
423 octahedral Fe that leads to visible wavelength surface scattering
424 and produces a spectrum that is broadly dark and flat with
425 Fresnel peaks near $0.65\ \mu\text{m}$ and a local reflectance minimum
426 near $0.55\ \mu\text{m}$ ^{49,50} (Figure 7). However, we note that Fe-sulfide
427 experiments with 100% Fe(II) (Figure 6D) have minor
428 hydration absorptions of $\sim 1.9\ \mu\text{m}$, reminiscent of the first
429 group and possibly an intermediate case that has produced
430 some amount of both magnetite and another hydrous phase.
431 The slight oxidation from our fully reduced Fe-sulfide minerals,
432 based on our colorimetry, may be a result of atmospheric
433 oxidation similar which will lead to a magnetite, which is a
434 stable mixed valent mineral under oxic conditions.
435

DISCUSSION

436
437 Organics present on Ceres are concentrated near Ernutet
438 crater, where the spectral bands ($3.3\text{--}3.5\ \mu\text{m}$) are prominent
439 and are likely of endogenous origin.¹ In proposed analogue
440 materials (CI, CM) to Ceres, organics (including amino acids)
441 are a sizable part of the bulk composition.²³ In addition to
442 organics, absorption bands have been interpreted as
443 ammoniated clays⁹ while other materials such as iron oxides
444 (magnetite) and sulfides are predicted to be present, based on
445 geochemical modeling.¹² Fe-hydroxide and Fe-sulfide minerals
446 are known for their ability to adsorb organic molecules onto
447 minerals.^{51–54} Yet, for every organic molecule tested, we did
448 not observe any condition in which a substantial amount of the
449 organic acids were adsorbed. At alkaline pH, we would expect
450 essentially all of the Fe in our experiments to be precipitated in
451 the solid phase, and considering that the Fe:organic ratio was
452 10:1, there was a large excess of mineral compared to organics
453 in these experiments. Therefore, an important finding from this
454 work is that in all conditions tested, most of the organics
455 remained in the aqueous phase, even in the presence of
456 minerals and therefore mobile/available for reactivity. Over the
457 2 weeks of sampling, we did not observe the concentration of
458 organics in the supernatant to change considerably (Figure 2).
459 This means that the adsorption capabilities of a given mineral/
460 organic combination are stable, at least in this experimental
461 time frame.

Organic Acid Adsorption onto Fe (Oxy)hydroxides.
462 Several factors can influence organic material adsorption on to 463

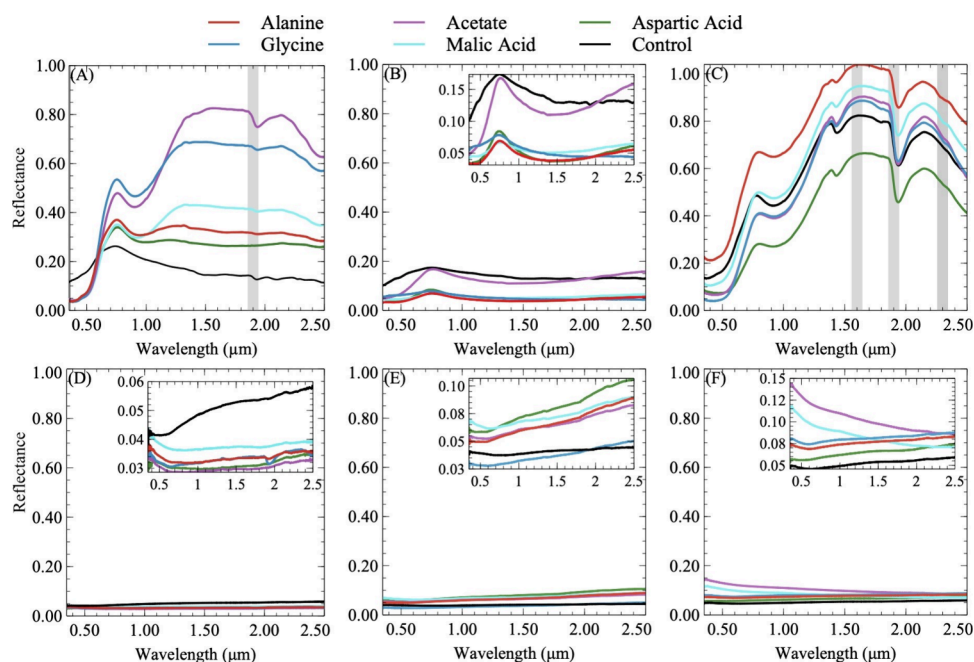


Figure 6. Spectral reflectance of Fe hydroxide and Fe-sulfide minerals reacted with and without organic acids. Reflectance spectra on the top show Fe-hydroxide experiments with and organic acids in experiments containing (A) 100% Fe(II)-hydroxides; the gray line at 1.9 μm indicates the hydration absorption feature (B) 50% = Fe(II)/Fe(III)-hydroxides (C) 100% Fe(III)-hydroxides; the gray lines at 1.6 and 1.9 μm indicate the hydration absorption feature, and the line at 2.3 μm indicates the metal–OH vibration. Reflectance spectra on the bottom show Fe-sulfide experiments containing (D) 100% Fe(II)-sulfides (E) 50% = Fe(II)/Fe(III)-sulfides and (F) 100% Fe(III)-sulfides.

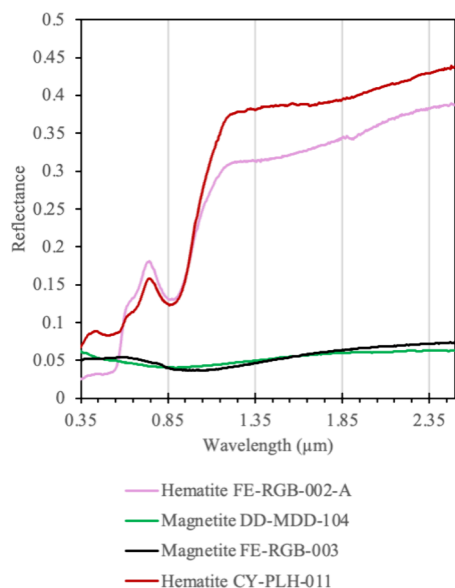


Figure 7. RELAB library VNIR reflectance spectra of hematite (pink, red) and magnetite (black, green)⁴² for comparison.

464 Fe-hydroxide minerals, for example, particle size, structure of
465 the mineral, solution composition, and type of adsorbate. An
466 adsorption study of arginine onto mesoporous materials
467 observed decreased adsorption as the ionic strength of the
468 experimental solution increased in experiments at pH 10.⁵⁸
469 Gao et al. explain three ionic strength factors NaCl content can
470 have on surface interactions: (1) the surface charge density of
471 the material becomes more negative; (2) NaCl negatively
472 affects the solubility of amino acids; (3) Na⁺ ions can block
473 negative charges on materials reducing adsorption capacity.⁵⁸
474 In studies using goethite (an Fe(III)-oxyhydroxide), oxalate,

malonate, phthalate, lactate, and citrate adsorption decreased
475 as pH increased.⁵⁹ Our experiments were initially titrated to
476 pH 10, meaning the surface of our materials would be
477 saturated with OH[−] groups, and the introduction of NaOH
478 into the solution might have influenced our experiments, as
479 Gao et al. explained.⁵⁸ Moreover, at this alkaline pH, the
480 organic material would become negatively charged species as
481 deprotonation occurs (pK_a's in Table 1), which would lead to
482 electrostatic repulsion onto the negatively charged mineral
483 (PZC in Table 2). In experiments by Schwaminger et al.,
484 12

Table 2. Surface Charge of Fe-Hydroxide Minerals and Fe-Sulfide Minerals

material	zero point charge (ZPC)	reference
green rust (sulfate and carbonate)	8.2	55
magnetite (Fe ₃ O ₄)	6.5	56
hematite (Fe ₂ O ₃)	8.6	56
FeS	3.3	57

amino acid adsorption onto nanomagnetite particles is
485 dependent on the side chain and point zero charge of the
486 adsorbent; the negatively charged carboxyl functional group is
487 likely the binding site on positively charged surfaces;³⁷
488 however, the functional groups in our organic acids may all
489 be deprotonated leading to more negative surface charges.
490

Organic Acid Adsorption onto Fe-Sulfides. Previously
491 reported density functional theory (DFT) adsorption studies
492 of adsorption of methylamine onto the FeS mineral
493 mackinawite observed adsorption on two surfaces of the
494 material.⁶⁰ In density functional theory studies of the amino
495 acid cysteine adsorbed onto FeS materials such as mackinawite,
496 the functional groups of cysteine (e.g., thiol, amine, and
497 carboxyl) were able to adsorb via H-bonding, van der Waals,
498

499 and ligand exchange mechanisms.⁶¹ While the mineral
500 generated in the study likely is not pure mackinawite, these
501 studies can contextualize what we are observing in our
502 experiments.

503 Previous work has shown that functional groups of our
504 organic material may also bind to mineral surface sites, for
505 example, glycine was able to bind in modeled simulations but
506 not in laboratory experiments.⁶² In the same review, alanine
507 and other amino acids were reported to weakly bind onto TiO₂
508 surfaces; alanine binds onto maghemite only by electrostatic
509 interactions.⁶² The organic materials we chose are negatively
510 charged at a pH of 10, and previous work has shown the pH
511 dependence on the binding of both mineral surfaces and
512 organic molecules. Work by Lambert on glycine adsorption on
513 silica surfaces suggests weak binding interactions.⁵² However,
514 for our minerals containing Fe, in several cases, the binding site
515 and strength are directly related to the zero-point charge
516 (ZPC) of the material. The ZPC of our materials is listed in
517 Table 2 and shows that our mineral surfaces are negatively
518 charged at a pH of 10 (Table 2). In addition to our
519 experimental conditions having both a negatively charged
520 mineral surface and negatively charged organic ion (negatively
521 charged ions > pK_a/pK₂), our experiments may be further
522 influenced by the solution's ionic strength.

523 In Fe-sulfide experiments, all the experiments beginning with
524 any Fe³⁺ showed Fe reduction to Fe²⁺ (and experiments
525 beginning with only Fe²⁺ remained reduced). Sulfide and Fe
526 are both redox active; sulfide oxidizes into various species,
527 potentially including elemental S or sulfate (SO₄²⁻), while our
528 Fe(III)-bearing minerals are reduced to ferrous precipitates.⁵³
529 Further, the conditions that simulated the Ceres brine
530 contained additional species (dissolved (NH₄)₂CO₃,
531 NaHCO₃, NH₄Cl, MgCl₂•6H₂O, NiCl₂, MgCO₃, KCl) that
532 increase the ionic strength. Our Ceres brine experiments have
533 an increased ionic strength in comparison to our experiments
534 without brine solution. The amount of Cl⁻ and Na⁺ ions is at
535 least 10 times the concentration of our amino acids in all of our
536 experiments. Previous research has demonstrated that high
537 ionic strength and pH play a critical role in decreasing
538 adsorption.^{58,62}

539 Since we observed that the organic material remains
540 primarily in solution (and not bound to the Fe-hydroxide or
541 sulfide minerals), the organics are mobile and available for
542 reactivity. Other researchers have performed reactions with
543 precursors, such as pyruvate, to the organic acids that were
544 studied in the work presented here.^{63,64} Possible organic
545 chemistry on Ceres would be relevant for understanding what
546 organic materials to expect and where they can be found on
547 Ceres for future missions. Further understanding the organic
548 inventory and its fate in the environment of Ceres is also
549 important to understanding habitability.

550 ■ CONCLUSIONS

551 Fe-hydroxides and Fe-sulfides may adsorb organic material, but
552 this is highly dependent on the mineral structure, pH, and
553 ionic strength of the solution under Ceres-relevant conditions.
554 In our experiments, we observed negligible adsorption of
555 organic material onto Fe-hydroxides and Fe-sulfides under
556 alkaline conditions. This finding suggests that the organic
557 material remained in the aqueous phase and may be easily
558 transported in Fe-hydroxide- and Fe-sulfide-rich environments.
559 While the alkaline pH of our experimental systems favors the
560 precipitation of iron, it also would create minerals with

negatively charged surfaces, which would repel the negatively
561 charged organic molecules. In the presence of Ceres analogue
562 brines, we observed little to no adsorption, even with dry/
563 wetting cycles. This work could imply good sites for Ceres
564 sample return concepts, as the organic material is indicated to
565 be mobile under these conditions. 566

■ ASSOCIATED CONTENT

SI Supporting Information

The Supporting Information is available free of charge at
569 [https://pubs.acs.org/doi/10.1021/acsearthspace-](https://pubs.acs.org/doi/10.1021/acsearthspacechem.4c00372)
570 [chem.4c00372.](https://pubs.acs.org/doi/10.1021/acsearthspacechem.4c00372) 571

Complete list of experimental conditions, reaction
572 images, ¹H NMR spectra of the reaction time points, 573
Ceres brine solution recipe, VNIR spectra, and color-
574 imetry data (PDF) 575

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617 Author Contributions

618 The manuscript was written through the contributions of all
619 authors. All authors have given approval to the final version of
620 the manuscript. E.M. contributed by writing the manuscript
621 and performing experiments. E.F. and D.V. performed
622 experiments. J.M.W. contributed by analyzing NMR. D.V.
623 performed NMR. R.Y.S. performed VNIR. R.P.H., J.C.-R.,
624 M.M.D., and B.L.H. contributed to the editing of the
625 manuscript. L.M.B. contributed by designing the experiments
626 and interpreting the data. All authors contributed to the editing
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642 Notes

643 The authors declare no competing financial interest.

644 ■ REFERENCES

- 645 (1) De Sanctis, M. C.; Ammannito, E.; McSween, H. Y.; Raponi, A.;
646 Marchi, S.; Capaccioni, F.; Capria, M. T.; Carrozzo, F. G.; Ciarniello,
647 M.; Fonte, S.; Formisano, M.; Frigeri, A.; Giardino, M.; Longobardo,
648 A.; Magno, G.; McFadden, L. A.; Palomba, E.; Pieters, C. M.; Tosi, F.;
649 Zambon, F.; Raymond, C. A.; Russell, C. Localized aliphatic organic
650 material on the surface of Ceres. *Science* **2017**, *355*, 719–722.
- 651 (2) De Sanctis, M. C.; Mitri, G.; Castillo-Rogez, J.; House, C. H.;
652 Marchi, S.; Raymond, C. A.; Sekine, Y. Relict Ocean Worlds: Ceres.
653 *Space Sci. Rev.* **2020**, *216*, 1–33.
- 654 (3) Russell, C. T.; Raymond, C. A.; Ammannito, E.; Buczkowski, D.
655 L.; De Sanctis, M. C.; Hiesinger, H.; Jaumann, R.; et al. Dawn arrives
656 at Ceres: Exploration of a small, volatile-rich world. *Science* **2016**, *353*,
657 1008–1010.
- 658 (4) Castillo-Rogez, J.; Brophy, J.; Miller, K.; Sori, M.; Scully, J.;
659 Quick, L.; Grimm, R.; Zolensky, M.; et al. Concepts for the future
660 exploration of dwarf planet Ceres' habitability. *Planet Sci. J.* **2022**, *3*,
661 41.
- 662 (5) Castillo-Rogez, J. Future exploration of Ceres as an ocean world.
663 *Nat. Astron.* **2020**, *4*, 732–734.
- 664 (6) National Academies of Sciences, Engineering, and Medicine.
665 *Origins, worlds, and life: a decadal strategy for planetary science and*
666 *astrobiology 2023–2032*. The National Academies Press: Washington
667 DC, 2023.
- 668 (7) Castillo-Rogez, J. C.; Neveu, M.; Scully, J. E. C.; House, C. H.;
669 Quick, L. C.; Bouquet, A.; Miller, K.; Bland, M.; et al. Ceres:
670 astrobiological target and possible ocean world. *Astrobio.* **2020**, *20*,
671 269–291.
- 672 (8) Hendrix, A. R.; Hurford, T. A.; Barge, L. M.; Bland, M. T.;
673 Bowman, J. S.; Brinckerhoff, W.; Buratti, B. J.; et al. The NASA
674 roadmap to ocean worlds. *Astrobio.* **2019**, *19*, 1–27.
- 675 (9) Ammannito, E.; De Sanctis, M.; Ciarniello, M.; Frigeri, A.;
676 Carrozzo, F. G.; Combe, J.-P.; Ehlmann, B.; Marchi, S.; McSween, H.
677 Y.; Raponi, A.; et al. Distribution of phyllosilicates on the surface of
678 Ceres. *Science* **2016**, *353*, No. aaf4279.
- 679 (10) De Sanctis, M. C.; Ammannito, E.; Raponi, A.; Marchi, S.;
680 McCord, T. B.; McSween, H. Y.; Capaccioni, F.; Capria, M. T.; et al.

- Ammoniated phyllosilicates with a likely outer Solar System origin on
(1) Ceres. *Nature* **2015**, *528*, 241–244. 682
- (11) Marchi, S.; Raponi, A.; Prettyman, T. H.; De Sanctis, M. C.;
Castillo-Rogez, J. C.; Raymond, C. A.; Ammannito, E.; Bowling, T.;
Ciarniello, M.; et al. An aqueously altered carbon-rich Ceres. *Nat.*
Astron. **2019**, *3*, 140–145. 686
- (12) Castillo-Rogez, J. C.; Neveu, M.; McSween, H. Y.; Fu, R. R.;
Toplis, M. J.; Prettyman, T. Insights into Ceres's evolution from
surface composition. *Meteorit. Planet. Sci.* **2018**, *53*, 1820–1843. 688
- (13) Marchi, S.; Raponi, A.; Prettyman, T. H.; De Sanctis, M. C.;
Castillo-Rogez, J.; Raymond, C. A.; et al. An aqueously altered
carbon-rich Ceres. *Nat. Astron.* **2019**, *3*, 140–145. 692
- (14) Bowling, T. J.; Johnson, B. C.; Marchi, S.; De Sanctis, M. C.;
Castillo-Rogez, J. C.; Raymond, C. A. An endogenic origin of cerean
organics. *Earth Planet. Sci. Lett.* **2020**, *534*, No. 116069. 695
- (15) McSween, H. Y., Jr.; Emery, J. P.; Rivkin, A. S.; Toplis, M. J.;
Castillo-Rogez, J. C.; Prettyman, T. H.; De Sanctis, C. M.; Pieters, C.
M.; Raymond, C. A.; Russell, C. T. Carbonaceous chondrites as
analogs for the composition and alteration of Ceres. *Meteorit. Planet.*
Sci. **2017**, *53*, 1793–1804. 700
- (16) Flores, E.; Martinez, E.; Rodriguez, L. E.; Weber, J. M.;
Khodayari, A.; VanderVelde, D. G.; Barge, L. M. Effects of amino
acid on phosphate adsorption onto iron (oxy) hydroxide minerals
under early earth conditions. *ACS Earth Space Chem.* **2021**, *5* (5),
1048–1057. 705
- (17) Bhattacharyya, A.; Schmidt, M. P.; Stavitski, E.; Azimzadeh, B.;
Martínez, C. E. Ligands representing important functional groups of
natural organic matter facilitate Fe redox transformations and
resulting binding environments. *Geochim. Cosmochim. Acta* **2019**,
251, 157–175. 710
- (18) Joshani, A.; Mirzaei, Y.; Barber, A.; Balind, K.; Gobeil, C.;
Gélinas, Y. Organic matter preservation through complexation with
iron minerals in two basins of a dimictic boreal lake with contrasting
deep water redox regimes. *Sci. Total Environ.* **2024**, *925*, No. 171776. 714
- (19) dos Santos, R.; Patel, M.; Cuadros, J.; Martins, Z. Influence of
mineralogy on the preservation of amino acids under simulated Mars
conditions. *Icarus* **2016**, *277*, 342–353. 717
- (20) De Sanctis, M. C.; Raponi, A.; Ammannito, E.; Ciarniello, M.;
Toplis, M. J.; McSween, H. Y.; Castillo-Rogez, J. C.; Ehlmann, B. L.;
Carrozzo, F. G.; Marchi, S.; Tosi, F.; Zambon, F.; Capaccioni, F.;
Capria, M. T.; Fonte, S.; Formisano, M.; Frigeri, A.; Giardino, M.;
Longobardo, A.; Magni, G.; Palomba, E.; McFadden, L. A.; Pieters, C.
M.; Jaumann, R.; Schenk, P.; Mugnuolo, R.; Raymond, C. A.; Russell,
C. T. Bright carbonate deposits as evidence of aqueous alteration on
(1) Ceres. *Nature* **2016**, *536* (7614), 54–57. 725
- (21) Durmus, Z.; Kavas, H.; Toprak, M. S.; Baykal, A.; Altınçekiç, T.
G.; Aslan, A.; Bozkurt, A.; Coşgun, S. L-lysine coated iron oxide
nanoparticles: synthesis, structural and conductivity characterization.
J. Alloys Compd. **2009**, *484*, 371–376. 729
- (22) Pušnik, K.; Peterlin, M.; Cigić, I. K.; Marolt, G.; Kogej, K.;
Mertelj, A.; Gyergyek, S.; Makovec, D. Adsorption of amino acids,
aspartic acid, and lysine onto iron-oxide nanoparticles. *J. Phys. Chem.*
C **2016**, *120*, 14372–14381. 733
- (23) De Sanctis, M. C.; Vinogradoff, V.; Raponi, A.; Ammannito, E.;
Ciarniello, M.; Carrozzo, F. G.; De Angelis, S.; Raymond, C. A.;
Russell, C. T. Characteristics of organic matter on Ceres from VIR/
Dawn high spatial resolution spectra. *Mon. Not. R. Astron. Soc.* **2019**,
482, 2407–2421. 738
- (24) Vinogradoff, V.; Poggiali, G.; Raponi, A.; Ciarniello, M.; De
Angelis, S.; Ferrari, M.; Castillo-Rogez, J. C.; Brucato, J.; De Sanctis,
M. C. Laboratory investigations coupled to VIR/Dawn observations
to quantify the large concentrations of organic matter on Ceres. *739*
Minerals. **2021**, *11*, 719. 743
- (25) Hoehler, T.; Bains, W.; Davila, A.; Parenteau, M.; Pohorille, A.
Life's requirements, habitability, and biological potential. *Planet.*
Astrobiol. **2020**, *13*, 37. 746
- (26) Eschenmoser, A. The search for the chemistry of life's origin.
Tetrahedron **2007**, *63* (52), 12821–12844. 748

- 749 (27) Williamson, M. P. Autocatalytic Selection as a Driver for the
750 Origin of Life. *Life* **2024**, *14* (5), 590.
- 751 (28) Miller, S. L. The mechanism of synthesis of amino acids by
752 electric discharges. *Biochim. Biophys. Acta* **1957**, *23*, 480–489.
- 753 (29) Barge, L. M.; Flores, E.; Baum, M. M.; VanderVelde, D. G.;
754 Russell, M. J. Redox and pH gradients drive amino acid synthesis in
755 iron oxyhydroxide mineral systems. *Proc. Natl. Acad. Sci. U.S.A.* **2019**,
756 *116*, 4828–4833.
- 757 (30) Koga, T.; Naraoka, H. Synthesis of amino acids from aldehydes
758 and ammonia: Implications for organic reactions in carbonaceous
759 chondrite parent bodies. *Earth Space Chem.* **2022**, *6*, 1311–1320.
- 760 (31) Otake, T.; Wesolowski, D. J.; Anovitz, L. M.; Allard, L. F.;
761 Ohmoto, H. Mechanisms of iron oxide transformations in hydro-
762 thermal systems. *Geochim. Cosmochim. Acta* **2010**, *74* (21), 6141–
763 6156.
- 764 (32) Peiffer, S.; Behrends, T.; Hellige, K.; Larese-Casanova, P.; Wan,
765 M.; Pollok, K. Pyrite formation and mineral transformation pathways
766 upon sulfidation of ferric hydroxides depend on mineral type and
767 sulfide concentration. *Chem. Geol.* **2015**, *400*, 44–55.
- 768 (33) Barge, L. M.; Flores, E.; Weber, J. M.; Fraeman, A. A.; Yung, Y.
769 L.; VanderVelde, D.; Martinez, E.; Castonguay, A.; Billings, K.; Baum,
770 M. M. Prebiotic reactions in a Mars analog iron mineral system:
771 effects of nitrate, nitrite, and ammonia on amino acid formation.
772 *Geochim. Cosmochim. Acta* **2022**, *336* (336), 469–479.
- 773 (34) Novikov, Y.; Copley, S. D. Reactivity landscape of pyruvate
774 under simulated hydrothermal vent conditions. *Proc. Natl. Acad. Sci.*
775 *U.S.A.* **2013**, *110*, 13283–13288.
- 776 (35) Huber, C.; Wächtershäuser, G. α -Hydroxy and α -amino acids
777 under possible Hadean, volcanic origin-of-life conditions. *Science*
778 **2006**, *314* (314), 630–632.
- 779 (36) Aguirre, V. P.; Jovic, S.; Webster, P.; Buser, C.; Moss, J. A.;
780 Barge, L. M.; Tang, Y.; Guo, Y.; Baum, M. M. Synthesis and
781 characterization of mixed-valent iron layered double hydroxides
782 (“green rust”). *Earth Space Chem.* **2021**, *5*, 40–54.
- 783 (37) Schwaminger, S. P.; García, P. F.; Merck, G. K.; Bodensteiner,
784 F. A.; Heissler, S.; Günther, S.; Berensmeier, S. Nature of interactions
785 of amino acids with bare magnetite nanoparticles. *J. Phys. Chem. C*
786 **2015**, *119*, 23032–23041.
- 787 (38) Locke, M. J.; McIver, R. T., Jr Effect of solvation on the acid/
788 base properties of glycine. *J. Am. Chem. Soc.* **1983**, *105*, 4226–4232.
- 789 (39) Reisinger, H.; King, C. J. Extraction and sorption of acetic acid
790 at pH above pKa to form calcium magnesium acetate. *Ind. Eng. Chem.*
791 *Res.* **1995**, *34*, 845–852.
- 792 (40) Max, J. J.; Chapados, C. Infrared spectroscopy of aqueous
793 carboxylic acids: malic acid. *J. Phys. Chem. A* **2002**, *106*, 6452–6461.
- 794 (41) Urry, D. W.; Gowda, D. C.; Peng, S.; Parker, T. M.; Jing, N.;
795 Harris, R. D. Nanometric design of extraordinary hydrophobic-
796 induced pKa shifts for aspartic acid: Relevance to protein
797 mechanisms. *Biopolymers: Original Research on Biomolecules.* **1994**,
798 *34*, 889–896.
- 799 (42) Clark, R. N.; Roush, T. L. Reflectance spectroscopy:
800 Quantitative analysis techniques for remote sensing applications. *J.*
801 *Geophys. Res.: Solid Earth.* **1984**, *89*, 6329–6340.
- 802 (43) Bishop, J. L.; Pieters, C.; Burns, R. G. Reflectance and
803 Mössbauer spectroscopy of ferrihydrite-montmorillonite assemblages
804 as Mars soil analog materials. *Geochim. Cosmochim. Acta* **1993**, *57*,
805 4583–4595.
- 806 (44) Horgan, B. H.; Cloutis, E. A.; Mann, P.; Bell, J. F., III Near-
807 infrared spectra of ferrous mineral mixtures and methods for their
808 identification in planetary surface spectra. *Icarus.* **2014**, *234*, 132–154.
- 809 (45) Viviano, C. E.; Seelos, F. P.; Murchie, S. L.; Kahn, E. G.; Seelos,
810 K. D.; Taylor, H. W.; Taylor, K.; et al. Revised CRISM spectral
811 parameters and summary products based on the currently detected
812 mineral diversity on Mars. *J. of Geophys. Res.: Planets.* **2014**, *119*,
813 1403–1431.
- 814 (46) Milliken, R. RELAB spectral library bundle. *NASA Planet. Data*
815 *Syst.* **2020**, 98.
- (47) Sklute, E. C.; Jensen, H. B.; Rogers, A. D.; Reeder, R. J. 816
Morphological, structural, and spectral characteristics of amorphous 817
iron sulfates. *J. Geophys. Res.: Planets* **2015**, *120*, 809–830. 818
- (48) Sklute, E. C.; Kashyap, S.; Dyar, M. D.; Holden, J. F.; Tague, 819
T.; Wang, P.; Jaret, S. J. Spectral and morphological characteristics of 820
synthetic nanophase iron (oxyhydr)oxides. *Phys. Chem. Miner.* **2018**, 821
45 (1), 1–26. 822
- (49) Sheppard, R. Y.; Milliken, R. E.; Russell, J. M.; Dyar, M. D.; 823
Sklute, E. C.; Vogel, H.; Melles, M.; et al. Characterization of iron in 824
Lake Towuti sediment. *Chem. Geol.* **2019**, *512*, 11–30. 825
- (50) Izawa, M. R.; Cloutis, E. A.; Rhind, T.; Mertzman, S. A.; 826
Applin, D. M.; Stromberg, J. M.; Sherman, D. M. Spectral reflectance 827
properties of magnetites: Implications for remote sensing. *Icarus.* **2019**, 828
319, 525–539. 829
- (51) Matrajt, G.; Blanot, D. Properties of synthetic ferrihydrite as an 830
amino acid adsorbent and a promoter of peptide bond formation. 831
Amino acids. **2004**, *26*, 153–158. 832
- (52) Lambert, J. F. Adsorption and polymerization of amino acids 833
on mineral surfaces: a review. *Orig. Life Evol. Biosph.* **2008**, *38*, 211– 834
242. 835
- (53) Firer, D.; Friedler, E.; Lahav, O. Control of sulfide in sewer 836
systems by dosage of iron salts: comparison between theoretical and 837
experimental results, and practical implications. *Sci. Total Environ.* 838
2008, *392*, 145–156. 839
- (54) Jonsson, C. M.; Jonsson, C. L.; Estrada, C.; Sverjensky, D. A.; 840
Cleaves, H. J., II; Hazen, R. M. Adsorption of l-aspartate to rutile (α - 841
TiO₂): experimental and theoretical surface complexation studies. 842
Geochim. Cosmochim. Acta **2010**, *74*, 2356–2367. 843
- (55) Guilbaud, R.; White, M. L.; Poulton, S. W. Surface charge and 844
growth of sulphate and carbonate green rust in aqueous media. 845
Geochim. Cosmochim. Acta **2013**, *108*, 141–153. 846
- (56) Xu, Y.; Schoonen, M. A. The absolute energy positions of 847
conduction and valence bands of selected semiconducting minerals. 848
Am. Mineral. **2000**, *85* (3–4), 543–556. 849
- (57) Bebie, J.; Schoonen, M. A. A.; Fuhrmann, M.; Strongin, D. R. 850
Surface charge development on transition metal sulfides: an 851
electrokinetic study. *Geochim. Cosmochim. Acta* **1998**, *62*, 633–642. 852
- (58) Gao, Q.; Xu, W.; Xu, Y.; Wu, D.; Sun, Y.; Deng, F.; Shen, W. 853
Amino acid adsorption on mesoporous materials: influence of types of 854
amino acids, modification of mesoporous materials, and solution 855
conditions. *J. Phys. Chem. B* **2008**, *112*, 2261–2267. 856
- (59) Filius, J. D.; Hiemstra, T.; Van Riemsdijk, W. H. Adsorption of 857
small weak organic acids on goethite: Modeling of mechanisms. *J.* 858
Colloid Interface Sci. **1997**, *195*, 368–380. 859
- (60) Dzade, N. Y.; Roldan, A.; De Leeuw, N. H. Adsorption of 860
methylamine on mackinawite (FeS) surfaces: A density functional 861
theory study. *J. Chem. Phys.* **2013**, *139*, No. 124708. 862
- (61) Dzade, N. Y.; Roldan, A.; de Leeuw, N. H. Surface and shape 863
modification of mackinawite (FeS) nanocrystals by cysteine 864
adsorption: a first-principles DFT-D2 study. *Phys. Chem. Chem.* 865
Phys. **2016**, *18*, 32007–32020. 866
- (62) Costa, D.; Savio, L.; Pradier, C. M. Adsorption of amino acids 867
and peptides on metal and oxide surfaces in water environment: a 868
synthetic and prospective review. *J. Phys. Chem. B* **2016**, *120*, 7039– 869
7052. 870
- (63) Vu, T. H.; Reynoso, L. R.; Johnson, P. V.; Hodyss, R. Amino 871
Acid-Mediated Formation of CO₂ in Flash-Frozen Ceres Brines. 872
Earth Space Chem. **2024**, *8*, 1214–1223. 873
- (64) Weber, J. M.; Czaplinski, E. C.; Henderson, B. L.; Barge, L. M.; 874
Castillo-Rogez, J. C.; Hodyss, R. Photochemical Stability and 875
Reactivity of Sodium Pyruvate: Implications for Organic Analysis 876
on Ceres. *ACS Earth Space Chem.* **2024**, *8*, 1385–1393. 877